Introduction

In order to evaluate the effect of a drug on the brain, it must successfully reach the brain. Systemic drug administration is the most commonly used method of drug administration. To reach the brain, drugs administered systemically must cross the blood-brain barrier. However, many drugs do not successfully cross the blood-brain barrier. Therefore, systemic drug administration may be an inadequate way to determine the effects of drugs on the brain. Consequently, in many cases, a topical injection administered directly into the brain is the optimal way to generate reliable data.

A brain infusion cannula connected to a mini-osmotic pump may be a good delivery method. Several studies have used brain infusion cannula as a delivery method.\(^1\)\(^2\) Fixation of the infusion cannula is mandatory for accurate delivery. Dental cements or adhesive gel have been used for fixation. However, these materials take time to harden and occasionally, fixation may fail. Therefore, we developed a novel method to fix the brain infusion cannula. Brain infusion cannulas were implanted into seven Wistar rats using the novel tie method. One, three, and seven days after the surgery, displacement of the brain infusion cannula and cerebrospinal fluid (CSF) leakage were evaluated in all the rats. None of the animals showed displacement of the brain infusion cannula or leakage of CSF at any point of time. In this study, we showed the successful results of the novel tie method for fixation of the brain infusion cannula, which needs no additional materials. It only requires a thread which is used for skin sutures.

Methods

This study was approved by the Institutional Animal Care and Use Committee of Chung-Ang University (2016-00086). A total of seven male Wistar rats (7-weeks old, 200–250 g)
were used. All animals were acclimatized to the laboratory conditions for 1 week before the start of the experiment. They were housed in a temperature- and humidity-controlled room with a 12 hour contrast cycle with free access to food and water.

**Surgical procedure**

The animals were anesthetized by intraperitoneal administration of ketamine hydrochloride (100 mg/kg; Ketamine, Yuhan, Seoul, Korea) mixed with xylazine (10 mg/kg; Rompun, Bayer-Korea, Seoul, Korea). The surgical procedure is shown in Fig. 1. First, the hair on the skull was shaved. After local anesthesia with 1% lidocaine hydrochloride, a curved incision was made on the skull. The skin flap was elevated and periosteum on the skull was exposed. Then, the periosteum was harvested from the skull and bregma was identified. A burr hole was made in the skull (AP: -0.3, L: 1.2, V: 4.5), according to the atlas of Paxinos and Watson (2006) to approach the right cerebral lateral ventricle. The temporalis muscles were sutured bilaterally using a non-absorbable monofilament [e.g., Blue Nylon 5-0 (Ailee, Busan, Korea)] but tie was not made. Harvested periosteum was place around the burr hole. A brain infusion cannula connected to a mini-osmotic pump was filled with 0.4% carmine in saline and implanted through the burr hole. Harvested periosteum was located beneath the brain infusion cannula. The non-absorbable monofilament was positioned on the brain infusion cannula, following which a tie was made tightly to allow no room between the brain infusion cannula, periosteum, and skull. Finally, the skin flap was repositioned and sutured.

**Evaluation of displacement of the brain infusion cannula and cerebrospinal fluid leakage**

One day after the surgery, all animals were anesthetized as described above. Skin flap on the skull was elevated again and position of the brain infusion cannula was checked. The area around the brain infusion cannula was inspected with the aid of a microscope for the appearance of fluid collection and blue pigmentation. Three and seven days after the surgery, the same procedure was repeated to evaluate the displacement of the brain infusion cannula and cerebrospinal fluid (CSF) leakage.

**Results**

No animal died during the procedure. One, three, and seven days after the surgery, displacement of the brain infusion cannula and CSF leakage were evaluated in the seven rats. There were no animals showing a change in the location of the brain infusion cannula or CSF leakage (Fig. 2).

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**Fig. 1.** Surgical procedure of the novel tie method. Elevation of the skin flap (A). Harvesting of the periosteum (B). Making a burr hole (C). Suturing temporalis muscles at both sides (D, E). Placement of the harvested periosteum around the burr hole (F). Implantation of the brain infusion cannula on the harvested periosteum (G). Making a tight tie to prevent room between the brain infusion cannula, periosteum, and skull (H).
Although brain infusion cannula is a good method to deliver compounds directly into the brain, fixation of the brain infusion cannula is essential for successful delivery. Failure of fixation may cause not only failure of delivery of the compounds but also CSF leakage. Dental cements and adhesive gel are the commonly used materials for the fixation of infusion cannula. However, since these materials take time to harden, the cannula need to be kept dry during this time for successful fixation. Thus, these methods require not only additional time but also cost more. Moreover, fixation may fail occasionally. Consequently, we developed a novel method, which needs no additional materials. The fixation requires only the thread used for skin sutures. Furthermore, this method does not need a dry field and there is no need to wait until it hardens. Therefore, we think that this method will help in making experiments that use brain infusion cannula more convenient and faster.

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